Elsevier Editorial System(tm) for Ultrasound

in Medicine and Biology

Manuscript Draft

Manuscript Number: UMB-D-16-00303R1

Title: Muscular echovariation: a new biomarker in Amyotrophic Lateral Sclerosis

Article Type: Original Contribution

Keywords: clinical neurology examination; observational study; ultrasound; amyotrophic lateral sclerosis.

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Abstract: The purpose is to assess the characteristics of echovariation in amyotrophic lateral sclerosis (ALS) compared with other MUS parameters. Twenty-six ALS patients (8 women, mean age 58.9 years, SD 12.02 yr) and 26 healthy controls (17 women; mean age 59.6 years, SD 6.41 yr) were included in this observational study. They underwent bilateral and transverse ultrasound of the biceps/brachialis, forearm flexor group, quadriceps femoris and tibialis anterior. Muscular thickness, echointensity and echovariation were analyzed. Muscles affected by ALS showed increased echointensity, decrease in thickness, and decrease echovariation. Echovariation in all muscles but quadriceps femoris, strongly correlated with muscle strength (explained variance between 21.8% in the biceps/brachialis and 37.5% in tibialis anterior) and the ALSFRS-R score (explained variance between 26% in the biceps/brachialis and 36.7% in the forearm flexor group). Echovariation is an easy to obtain QMUS parameter that could distinguish ALS from healthy controls more accurately than previous described biomarkers.

Suggested Reviewers:

Opposed Reviewers:

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1 ABSTRACT

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The purpose is to assess the characteristics of echovariation in amyotrophic lateral 3 4 sclerosis (ALS) compared with other MUS parameters. Twenty-six ALS patients (8 women, mean age 58.9 years, SD 12.02 yr) and 26 healthy controls (17 women; mean age 5 59.6 years, SD 6.41 yr) were included in this observational study. They underwent 6 bilateral and transverse ultrasound of the biceps/brachialis, forearm flexor group, 7 8 quadriceps femoris and tibialis anterior. Muscular thickness, echointensity and echovariation were analyzed. Muscles affected by ALS showed increased echointensity, 9 decrease in thickness, and decrease echovariation. Echovariation in all muscles but 10 quadriceps femoris, strongly correlated with muscle strength (explained variance between 11 21.8% in the biceps/brachialis and 37.5% in tibialis anterior) and the ALSFRS- R score 12 (explained variance between 26% in the biceps/brachialis and 36.7% in the forearm flexor 13 group). Echovariation is an easy to obtain QMUS parameter that could distinguish ALS 14 from healthy controls more accurately than previous described biomarkers. 15

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- -, 18

Keywords: clinical neurology examination; observational study; ultrasound; amyotrophic
lateral sclerosis.

INTRODUCTION

Amvotrophic lateral sclerosis (ALS) is a rapidly progressive neurodegenerative 3 disease involving the upper and lower motor neuron (LMN) that produces muscle 4 fasciculations, weakness and atrophy. Clinical monitoring of LMN is usually based on 5 testing muscular strength with the Medical Research Council scale (MRC) (Florence et 6 7 al. 1992) and the revised ALS functional rating scale, (ALSFRS-R) (Cedarbaum et al. 1999). However, these tools show limited sensitivity for measuring changes and, 8 9 consequently, clinical endpoints usually require prolonged follow up (Simon et al. 2014; Turner and Benatar 2015). In clinical trials, therefore, reliable biomarkers measurable 10 over the short term are desirable (Turner and Benatar 2015). 11

Although several neurophysiological tools have been developed as progression
biomarkers in ALS (Simon et al. 2014), they require wide operator experience, are time
consuming and pain is a considerable limiting factor (Simon et al. 2014).

Current ALS diagnostic criteria require the detection by electromyography (EMG) 15 of denervation signs or fasciculations in muscles with neurogenic changes (Costa et al. 16 2012). A fasciculation is an involuntary synchronous contraction of all the skeletal 17 muscle fibers within a single motor unit, which arises as a result of spontaneous 18 depolarization of a lower motor neuron. Fasciculation potentials of abnormal morphology 19 in the EMG recording are a characteristic clinical feature of ALS, but fasciculation 20 potentials of normal morphology can occur in healthy subjects (Brooks et al. 2000). 21 Muscle ultrasonography (MUS) is an accessible, painless and easy to perform 22 method to detect fasciculations and structural muscle changes in ALS (Arts et al. 2011a; 23 Simon et al. 2014). MUS has been shown more sensitive than EMG for detecting 24 fasciculations, especially in the bulbar region (Grimm et al. 2015; Misawa et al. 2011) 25 and structural changes such as decrease of muscle thickness and increase of echointensity 26 (EI) have been found (Arts et al. 2011a; Misawa et al. 2011; Simon et al. 2014). 27

However, their role as diagnostic or progression biomarkers is limited, because of the 1 high interindividual variability in structural parameters (Arts et al. 2011a; Misawa et al. 2 2011; Pillen et al. 2008). Therefore, it would be a major advance to find a MUS 3 4 biomarker not influenced by other factors than disease ones. Echovariation (EV) has been 5 previously established to characterize plantar fasciitis and it is a reproducible, short and 6 easy to carry out procedure (Ríos-Díaz et al. 2015). 7 We hypothesize that EV could act as a more reliable biomarker in ALS since it is an adimensional parameter (Arts et al. 2011a). Therefore, we compared muscle thickness, 8 9 EI and EV in four muscle groups in ALS patients and age matched controls and assessed how time and other clinical variables influenced these parameters. 10 11 12 **MATERIAL AND METHOD** 13 14 **Subjects selection** 15 16 Patients (n=26) were recruited from the Valencia ALS Association (ADELA) 17 between September 2013 and April 2014. The patients were diagnosed as having ALS by 18 an experienced neurologist (JFVC), according to the revised El Escorial Criteria (Brooks 19 20 et al. 2000). Twenty-six healthy volunteers without a history of hereditary neuromuscular 21 disease were recruited as control group. 22 23 24 Standard protocol approvals, registrations, and patient consents 25 26 This study was approved by the ethics committee of the Universidad Católica de 27 Murcia (Spain) and performed following the Helsinki Declaration principles. All 28 participants provided written informed consent. 29 30 31 **Recorded clinical variables** 32 33

Demographical and clinical characteristics (sex, age, weight, height, body mass index -BMI-, time of evolution from symptoms onset and date of diagnosis) were recorded. ALSFRS-R score (Cartwright et al. 2011) and muscle strength, measured with the Medical Research Council (MRC) rating scale (Arts et al. 2011a), were assessed by the same researcher (JMP) on the same day as the MUS was performed.

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7 Ultrasonography

MUS was performed in four muscle groups of each side in patients and controls 9 by the same experienced examiner (JMP) with the participants sitting and completely 10 relaxed. A LEbt12 phased array real-time scanner (software 2014) from General Electric 11 Company with a 5–13 MHz linear array transducer (12L-RS) was used for MUS. All 12 system-setting parameters, such as gain (98 dB), time gain compensation (in neutral 13 14 position), depth (5 cm for tibialis anterior and 6 cm for the other muscle groups), 15 frequency (12 Mhz), compression and focus (two focal points at 1.8 and 2.6 cm) were 16 kept constant throughout the study.

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To avoid oblique scanning angles the angle of the probe was adjusted until the best muscle EI was obtained in every image.

Applying the standardized protocol described by Arts et al. (2008) bilateral transverse ultrasound images of the biceps/brachialis group (2/3 distance acromion – antecubital crease), forearm flexors group (2/5 distance antecubital crease – distal end radius), quadriceps (1/2 distance anterior superior iliac spine – superior aspect patella) and tibialis anterior (1/4 distance inferior aspect patella – lateral malleolus) were obtained and measured. Three images were taken of every muscle in order to minimize variation in muscle thickness, EI and EV.

The resulting bitmaps had a resolution of 820 x 614 pixels (7.6 px/mm) with 256 grey levels and were stored as .TIFF files without compression or losses (Wiggins et al.

1 <mark>2001</mark>).

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Image analysis

Fasciculations as well as quantitative MUS variables (QMUS), including muscle
thickness, EI and EV, were obtained for each muscle group.

Muscle thickness was measured with electronic calipers as previously described 8 9 (Arts et al. 2010). The thickness of the biceps/brachialis group was measured between the uppermost part of the bone echo of the humerus and the superficial fascia of the biceps; 10 11 the forearm flexor group between the interosseous membrane (next to the radius) and the superficial fascia of the most ventral flexors; the quadriceps femoris between the 12 13 uppermost part of the bone echo of the femur and the superficial fascia of the rectus 14 femoris (which includes the vastus intermedius); and the tibialis anterior between the interosseous membrane (next to the tibia) and the ventral fascia of the tibialis anterior 15 (Figure 1). 16

17 Fasciculations were registered in each muscle for 10 seconds as previously18 published (Arts et al. 2008).

Thickness was measured in all three images of each muscle group by an expert
ultrasonographist (JMP) and the mean of the three values was used for the corresponding
analysis.

Image processing and analysis was performed by one researcher (JRD) using the ImageJ (v.1.48) software. This researcher, who was blind for diagnosis, selected the region of interest (ROI) the ROI Manager application for ImageJ, with a size of 71 x 40 pixels for tibialis anterior and 73 x 73 pixels for other muscle groups on an 8-bit gray scale. The ROI was defined as the muscle region without bone and fascia with the best reflection (Figure 1).

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EV, a parameter that can be interpreted as a measure of intensity range (Ríos-

1	Díaz et al. 2015), was determined by the relation between standard deviation and mean of
2	pixel intensity obtained from the histogram:
3	$EV = \sigma/\mu \cdot 100$
4 5	where σ is the standard deviation of the image intensities and μ is the mean value of
6	intensity in each ROI.
7	EI and EV were obtained from the ROIs of three images of each muscle and the
8	mean of the three values was used for analysis. Sets of 20 images for each muscular
9	group were analyzed by another researcher (MEDBA) who was blinded to the previous
10	results to analyze inter-observer reliability in thickness measurement and ROI selection.
11	EI and EV of each ROI and muscle thickness were calculated.
12 13	Statistics
13 14	Statistics
15	Data were analyzed using IBM SPSS Statistics for Windows 19.0 (IBM
16	Company, 2010).
17	Variables were checked for normality by the Kolmogorov-Smirnov test and for
18	the homogeneity of variances by the Levene test. In addition, we analyzed the asymmetry
19	and kurtosis coefficients and the normality Q-Q plots.
20	Data were summarized as mean and standard deviations (SD), with 95%
21	confidence intervals (CI) for continuous variables, and absolute and relative frequencies
22	for categorical variables.
23	Intraclass correlation coefficient (ICC) for 2-way mixed effect model and absolute
24	agreement was calculated to determine inter-observer reliability of thickness measure, EI
25	and EV. Next criteria were used to judge the reliability coefficients: very low (<0.20),
26	low (0.21-0.40), moderate (0.41-0.60), good (0.61-0.80) and very good (0.81-1.00).
27	Unpaired t-tests were used to compare continuous variables and chi- square test to
28	compare categorical variables at baseline between ALS patients and controls.
29	Paired t-tests were used to assess right-left differences in muscle thickness, EI and

1 EV measurements (Armitage et al. 2002).

2 3 4 5	QMUS variables in patients and controls One-way ANCOVA was used to compare OMUS variables in patients and
6	controls, controlling for effects of clinical and demographical covariates (Feinstein 2002)
7	Cohen's d statistic was used to evaluate the effect sizes, in this case by dividing
8	mean score differences between patient and control group by the initial standard deviation
9	of the control group (Cohen 1988; Kelley and Preacher 2012). A Cohen's d statistic of
10	<0.1 corresponds to a small size effect, around 0.3 to a medium size effect and >0.5 a
11	large size effect.
12	Influence of time of evolution.
13 14	Regression models were used to study the relationship between time of evolution
15	(logarithmic transform was applied to this variable to correct the absence of a normal
16	distribution) as independent variable and the QMUS parameters as dependent variables
17	(Kleinbaum et al. 1998).
18 19	Relations between QMUS parameters and MRC- ALSFRS-R
20	MRC and ALSFRS-R were taken as dependent variables to check the relationship
21	with QMUS parameters (thickness, echointensity and echovariation as independent
22	variables).
23	All regressions were carried out with a fixed inclusion of ultrasonographics
24	parameters to obtain a raw model and the stepwise inclusion of sex, age, and BMI. The P-
25	in and P-out values were 0.05 and 0.10, respectively. To control for possible collinearity,
26	the tolerance to enter in the model was fixed at 0.01 (Hair et al. 2010).
27	The presence of influential observations was checked with the Cook distance (any
28	influential observation was considered a Cook distance of >1). Collinearity for
29	independent variables was evaluated with the tolerance and variance inflation factors
30	(Kleinbaum et al. 1998). Data are presented as b coefficient and 95% CI. The relation

1	between variables was studied with a partial correlation coefficient that adjusts the linear
2	relation between the dependent and independent variables. In addition, we calculated the
3	goodness of fit with the partial determination coefficient (r-squared in %).
4 5 6	P values of < 0.05 were considered statistically significant for all the tests.
7	RESULTS
8 9	Study subjects characteristics
10 11	Twenty-six participants with ALS (8 women, mean age 58.9 years, SD 12.02 yr)
12	and 26 healthy controls (17 women; mean age 59.6 years, SD 6.41 yr) were included in
13	this study. No significant differences in age, height and weight were found. BMI was
14	slightly different and sex distribution was significantly different. The clinical
15	characteristics along with their mean and standard deviation (SD) are shown in Table 1.
16 17 18 19	Ultrasound variables
20	Fasciculations were detected in 15 of the 26 patients (57.7%) and none of the
21	healthy controls. Altogether, they were detected in 30 out of 52 (57.7%) of the
22	biceps/brachialis group, 26 (50%) of the forearm flexor group, 22 (42.3%) of the
23	quadriceps femoris and 13 (25.7%) of tibialis anterior.
24	QMUS variables for each muscle and group are shown in Table 2. There were no
25	significant right-left differences in thickness, EI or EV in the four studied muscle groups,
26	so only one sample of each right/left muscle group was selected for further analysis (52
27	ultrasonograms for each group).
28	The inter-observer reliability ICCs for thickness measurement were over 0.97 for
29	all the muscular groups that revealed a very good reliability. Inter-observer measures of
30	echointensity revealed very good reliability too; the highest ICC was obtained for
31	quadriceps femoris (ICC=0.98; 95% CI= 0.95 to 0.99) and the lowest for tibialis anterior
32	(ICC=0.95; 95% CI= 0.87 to 0.98). As regards the reliability of echovariation, the inter- 9

1 observer ICCs were over 0.80 for all the muscular groups.

2	When analyzing QMUS differences between patients and controls, mean
3	comparisons were made with the corresponding corrections for sex and BMI (see
4	footnotes in tables for details). As expected, muscle thickness in the ALS group was
5	significantly lower than in the healthy controls for all analyzed muscles groups. EI was
6	higher in patients than in controls, except in the case of quadriceps femoris where no
7	differences between patients and controls were found. Finally, EV was significantly
8	lower in patients for all muscles, with a strong size effect (higher than 1.0), except for
9	quadriceps femoris which showed a moderate size effect of 0.55.
10 11 12 13	Relation between QMUS parameters and time from symptoms onset. Table 3 shows the results of the raw and adjusted regression analysis for age, sex
14	and BMI, when necessary.
15	No significant relationship was found between time of evolution and QMUS for
16	any muscle group, although in the biceps/brachialis group a 6.71% of the EV variance
17	could be explained by time of evolution, which was nearly significant.
18	BMI, age and sex showed an interaction with muscle thickness and EI, but not
19	with EV.
20 21	Relation between QMUS parameters and MRC-ALFSRS-R
22 23	Table 4 shows the relationships between the QMUS parameters and MRC ad
24	ALFSRS-R scores. Correlations are expressed as partial correlations, which provide
25	information on the explained variance about the individual variable without the effect of
26	the others.
27	A significant and positive relationship was found between thickness and EV, and
28	the MRC score in all regions, i.e. the greater the thickness or EV the higher the MRC
29	score. The explained variance for thickness was between 24.8% in the biceps/brachialis

group and 7.4% in the forearm flexor group. The explained variance for EV was between 1 26.5% in the tibialis anterior and 4.5% in the quadriceps femoris. The EI showed no 2 significant correlation with MRC in any muscular group. 3

4 Analysis of the relationship between the ALSFRS-R score and QMUS parameters showed similar results. Thickness and EV were significantly and directly correlated with 5 6 the ALSFRS-R score: the greater the thickness or EV the higher the ALSFRS-R score. 7 The explained variance for thickness was between 26.9% in the biceps/brachialis group and 7.5% in the forearm flexor group. The explained variance for EV was between 36.7% 8 9 in the forearm flexor group and 8.9% in the quadriceps femoris.

The EI showed an inverse correlation with the ALSFRS-R score only in tibialis 10 anterior (4.0% of explained variance). 11

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DISCUSSION 14

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We describe a new QMUS parameter (EV) that could be useful as diagnostic 16 and/or progression biomarker in ALS, and compared its characteristics with those of 17 18 previously described OMUS parameters (muscle thickness and EI).

Previous studies have shown that muscle thickness decreases and EI increases 19 even in clinically unaffected muscles (Arts et al. 2011a; Arts et al. 2012; Grimm et al. 20 2015), reflecting the progressive muscular atrophy and fibrosis that occurs in ALS as a 21 result of muscle denervation (Pillen et al. 2009). Recently, MUS has been proposed to 22 increase the sensitivity of EMG if both the presence of fasciculations and EI are taken 23 into account (Arts et al. 2012; Grimm et al. 2015), suggesting their role as diagnostic 24 biomarker. However, EMG remains necessary for ALS diagnosis, since decreased muscle 25 thickness and increased EI are age-dependent, show high interindividual variability, and 26 can also be observed in other neuromuscular diseases (Pillen et al. 2008). 27

The major advantage of MUS is that it is a painless technique. Consequently, its 28

1	role in monitoring progression in clinical trials could be of great value to avoid the
2	repetition of painful techniques such as EMG. However, correlation between EI and
3	muscle thickness and muscle strength and disability is only weak (Arts et al. 2011a;
4	Grimm et al. 2015) and disease progression measured as changes in MRC or ALSFRS- R
5	does not correlate with any of them (Arts et al. 2011a). The ultimate cause of this remains
6	unknown but suggests that factors other than disease related ones could influence changes
7	in EI or muscle thickness. Another reason may be that the reproducibility of these
8	measurement techniques is limited because of the ROI selection or because very
9	important adjustments such as frequency and depth of ultrasonography are not taken into
10	account or two different US devices are used (Arts et al. 2011b; Florence et al. 1992;
11	Pillen et al. 2009). Whatever the reason, it implies a strong limitation and a major
12	challenge of MUS when used as a progression biomarker.
10	Second order statistical methods of texture analysis have also been proposed to improve
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 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 	 accuracy and precision of these measures (Molinari et al. 2015). However, in comparison to EI or muscle thickness they require more complex post-processing operations, Consequently, reproducibility supposes a strong limitation and a major challenge of MUS when used as a progression biomarker. To minimize this risk, we performed all studies with the same US device, we strictly defined the acquisition parameter and the mean of three images of each muscle was calculated for each parameter. Moreover, the inter-observer reliability in ROI selection for EI and EV and thickness measure were vey good, suggesting a high reproducibility of our results. Fasciculations As expected, MUS identified frequent and widespread fasciculations. However, they were less frequent than previously reported (Grimm et al. 2015; Misawa et al. 2011),

moderate to advanced stage of the disease. Fasciculations are an early finding in ALS and
their frequency diminishes with time (de Carvalho and Swash 2013), which might explain
the relatively low frequency of fasciculations in our cohort, especially where the disease
started.

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Muscle thickness

As described in previous studies (Arts et al. 2008; Arts et al. 2012; Grimm et al.
2015), a decrease in muscle thickness was observed in ALS patients compared to healthy
controls. The highest size effect of this difference was obtained for quadriceps femoris.
As mentioned above, this finding could be related to the fact that in more than 50% of
subjects with ALS in our sample, symptoms started in lower limbs.

As previously published, muscle strength and disability showed a weak correlation with muscle thickness (Arts et al. 2011a) except for the quadriceps femoris where this correlation was moderate. No clear correlation was found between EV and disease duration.

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20 Echointensity

EI can be estimated using either a subjective visual grading scale (Grimm et al. 22 2015) or a quantitative methodology with an average gray-scale analysis that provides 23 more objective and potentially more sensitive information, but which does not contain 24 25 any data about tissue homogeneity (Pillen et al. 2008). Quantitative analysis of the EI is dependent on settings, the ultrasound device and on ROI selection, so that to record EI, a 26 standardized protocol and some degree of experience are necessary. New methods of 27 computer-assisted texture analysis, such as gray level co-occurrence matrix (Pillen et al. 28 2008; Ríos-Díaz et al. 2010) and fractal analysis (Gdynia et al. 2009), have been proposed 29 to quantify muscle alterations in neuromuscular disorders. However, these procedures 30

require expert handling of image analysis software and involve extra time for exportation
 and subsequent computer analysis, which could hinder their application in clinical
 practice.

4 In the present study we analyzed EI using the Arts protocol (Arts et al. 2008), but system-setting parameters such as frequency, gain, compression, time gain compensation, 5 6 focus (number and position) and depth were kept constant throughout our study to ensure 7 measurement reliability (Mayans et al. 2012). As suggested by Gdynia et al. (2009), we selected a small ROI, unlike other authors who selected the largest ROI possible (Arts et 8 al. 2008; Arts et al. 2011a; Arts et al. 2011b). In this last approach, large muscle areas are 9 evaluated, combining areas of maximum reflection with anisotropic areas, which cause a 10 decrease in EI. In our study, a ROI of the most reflective muscle segment, avoiding 11 anisotropic muscle areas was obtained. This ensures that zones are selected in which 12 muscle tissue presents maximum brightness. 13

EI was found to increase in all the muscles studied of ALS patients except, 14 intriguingly, the quadriceps femoris. Considering that symptoms in most patients started 15 in the lower limbs and that EI significantly interacted with sex, age and BMI, we suggest 16 17 that EI (at least in quadriceps femoris) can be influenced by non-disease related factors, limiting its usefulness. Moreover, as previously reported (Grimm et al. 2015), we found 18 no significant correlation between EI and time of evolution. Likewise, no correlation was 19 20 found with muscle strength or disability. Others have found a weak correlation with these variables (Arts et al. 2011a; Grimm et al. 2015); however, our statistical analysis (partial 21 correlations) was stronger than those, to avoid confounding variables. 22

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25

24 Echovariation

We propose a new MUS biomarker, EV, which is a first order statistical measure that quantifies the deviation of the level of gray from the average. It is a fast and easy method to obtain information on tissue homogeneity (Aggarwal and Agrawal 2012) and 14 1 an adimensional parameter, reducing the chance of the heterogeneity.

In our study, we observed a significantly lower level of EV in the muscles of ALS patients compared to healthy controls, with higher effect sizes than those found for muscle thickness or EI except for the quadriceps femoris, in which muscle thickness showed greater effect. Unlike EI, EV did not interacted with non-disease related factors such as age, sex or BMI, further suggesting its potential as biomarker. Moreover, muscle strength and disability strongly correlated with EV except, once again, for the quadriceps femoris. Conversely, no clear correlation was found between EV and disease duration.

9 There are some limitations in our study. First, the study is transversal and the 10 number of studied subjects was limited. Moreover, our cohort of ALS patients was in a 11 moderate to advanced disease stage and no ALS mimics subjects were studied. Therefore, 12 in order to establish the role of EV as a diagnostic, progression or prognostic biomarker, 13 these results must be replicated in a larger, prospective and longitudinal cohort of 14 suspected ALS patients.

In conclusion, we describe a new, easy to obtain QMUS parameter that seems to distinguish ALS from healthy controls more accurately than previous described biomarkers. It also seems to correlate better with strength and disability, limiting the influence of non-disease related factors.

19 20

21 Acknowledgments

22

This study was supported by the research grant from the Universidad Católica de
Murcia (PMAFI/10/14) Spain.

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FIGURE LEGENDS

Figure 1. The ultrasound measurement of echointensity and muscle thickness.

4 Ultrasonographic scans of the biceps/brachialis group (A-B), forearm flexor group (C-

5 D), quadriceps (E-F), and tibialis anterior (G-H). The left panel depicts muscle thickness

6 measured through electronic calipers in healthy subjects and the right panel represents

7 the region of interest for echointensity in subjects with ALS using the ImageJ (v.1.48)

8 software.

9 **Figure 2.** Comparison of quantitative muscular US parameters in controls and ALS patients.

10 Mean and 95% confidence interval for BBr (biceps/brachialis group), FFG (forearm flexors

11 group), QF (quadriceps femoris) and TB (tibialis anterior). * p<0.005; **p≤0.001. n.s=non-

12 significative.

Baseline characteristics	ALS Patients (n=26)	Healthy (n=26)	p-value
Females (n) (%)	8 (30.8 %)	17 (65.4 %)	< 0.001
Age (years)	58.9 (12.02); 55.8 to 62.0	59.6 (6.41); 57.9 to 61.4	0.570
Weight (kg)	69.9 (17.42); 65.4 to 74.4	72.4 (17.19); 67.6 to 77.2	0.154
Height (m)	1.67 (0.086); 1.65 to 1.69	1.66 (0.08); 1.63 to 1.68	0.773
BMI (kg/m^2)	24.9 (5.13); 23.6 to 26.3	26.2 (4.87); 24.9 to 27.6	0.050
Time from symptoms onset (months)	26.1 (15.77); 21.72 to 30.5		
Symptoms onset-diagnosis (months)	16.3 (9.89); 13.5 to 19.1		
Disease onset (n) (%)			
Right Lower Limb	9 (34.6 %)		
Left Lower Limb	5 (19.2 %)		
Right Upper Limb	1 (3.8 %)		
Left Upper Limb	4 (15.4 %)		
Bulbar	7 (26.9 %)		
ALSFRS-r (max 48)	26.2 (11.67); 22.9 to 29.4		
MRC (max 100)	58.5 (24.75); 51.7 to 65.4		

Table 1. Baseline characteristics of the study sample.
 1

2 3 4 5 6 Data are presented as mean (Standard deviation) and 95% of confidence interval for quantitative variables and as absolute frequencies (relative frequencies). BMI: Body Mass Index. ALSFRS-r: Amyotrophic lateral sclerosis

functional rating scale. MRC: Medical Research Council Scale for muscular Strength. P-value for Chi-square

test for sex differences and T-Student test for independent samples for age, weight, height and body mass index

differences.

QMUS	Patients (n=52)		Control		Size		
parameters	Mean (SD)	95% C.I.	Mean (SD) 95% C.I.		p-value	effect	
		Biceps/brack	/brachialis group ‡				
Thickness	28.6 (6.34)	26.8 to 30.3	32.8 (6.25)	31.1 to 34.6	< 0.001	0.90	
Echointensity	92.2 (14.4)	88.2 to 96.2	86.7 (8.78)	84.2 to 89.1	0.001	0.67	
Echovariation	23 (7.33)	21 to 25	29.2 (4.24)	28 to 30.4	< 0.001	1.08	
		Forearm fle	xor group ‡‡				
Thickness	30 (9.69)	27.3 to 32.7	31.3 (5.96)	29.7 to 33	0.016	0.42	
Echointensity	101.4 (15.25)	97.2 to 105.7	90.7 (15.07)	86.5 to 94.9	< 0.001	0.88	
Echovariation 19.3 (4.55)		18.1 to 20.6	18.1 to 20.6 25.5 (4.22)		< 0.001	1.09	
Quadriceps femoris §							
Thickness	22.9 (8.97)	20.4 to 25.4	29.4 (6.06)	27.7 to 31.1	< 0.001	1.00	
Echointensity	100.6 (18.03)	95.5 to 105.6	97 (12.77)	93.4 to 100.5	0.245	0.23	
Echovariation 18.9 (4.46)		17.6 to 20.1	21.7 (5.66)	20.2 to 23.3	0.005	0.55	
Tibialis a			nterior §§				
Thickness	19.1 (5.59)	17.5 to 20.6	21.7 (4.91)	20.3 to 23	< 0.001	0.91	
Echointensity	116 (16.36)	111.5 to 120.6	105.1 (14.63)	101.1 to 109.2	< 0.001	1.03	
Echovariation	16.5 (4.31)	15.3 to 17.7	25 (4.85)	23.6 to 26.3	< 0.001	1.35	

Table 2. Quantitative muscular differences between controls and ALS patients.

2 SD: Standard Deviation. CI 95%.: Confidence Interval. p-value for independent-samples T-Student test. ‡

3 Thickness ANCOVA corrected by sex and BMI, Echointensity and Echovariation corrected for sex. ‡‡

4 Thickness ANCOVA corrected for sex and BMI, Echointensity corrected for sex and Echovariation

5 corrected for BMI. § Thickness ANCOVA corrected for sex. §§ Thickness and Echointensity ANCOVA

6 corrected for sex. † Effect size was estimated with Cohen's d (low<0.2; moderate=0.5; large>0.80).

Raw				Adjusted				
QMUS	B coefficient (SE)	95% CI for B	p-value	Rp (Rp-squared %)*	B coefficient (SE)	95% CI for B	p-value	Rp (Rp-squared %)
Biceps/brachialis g	roup							
Thickness	-0.045 (0.057)	-0.16 to 0.070	0.431	-0.112 (1.25%)	-0.007 (0.047)	-0.10 to 0.09	0.881	-0.022 (0.05%)
Echointensity	-0.050 (0.129)	-0.31 to 0.21	0.702	-0.054 (0.29%)	-0.133 (0.118)	-0.37 to 0.10	0.263	-0.161 (2.6%)
Echovariation	-0.12 (0.064)	-0.25 to 0.010	0.064	-0.259 (6.71%)				
Forearm flexor gro	oup							
Thickness	-0.146 (0.084)	-0.315 to 0.02	0.091	-0.237 (5.61%)	-0.115 (0.072)	-0.26 to 0.03	0.116	-0.225 (5.08%)
Echointensity	-0.139 (0.135)	-0.411 to 0.13	0.308	-0.144 (2.07%)	-0.183 (0.133)	-0.45 to 0.08	0.175	-0.193 (3.73%)
Echovariation	-0.002 (0.041)	-0.084 to 0.08	0.954	-0.008 (0.01%)				
Quadriceps femori.	S							
Thickness	-0.052 (0.08)	-0.213 to 0.11	0.519	-0.092 (0.84%)	-0.015 (0.075)	-0.17 to 0.14	0.844	-0.028 (0.08%)
Echointensity	-0.218 (0.159)	-0.536 to 0.1	0.177	-0.19 (3.62%)				
Echovariation	0.003 (0.04)	-0.165 to 0.14	0.949	0.009 (0.01%)				
Tibialis anterior								
Thickness	0.01 (0.05)	-0.091 to 0.11	0.841	0.029 (0.08%)	0.042 (0.042)	-0.04 to 0.13	0.319	0.142 (2.02%)
Echointensity	0.004 (0.147)	-0.291 to 0.3	0.980	0.004 (0%)	-0.089 (0.125)	-0.34 to 0.16	0.480	-0.101 (1.02%)
Echovariation	0.011 (0.039)	-0.066 to 0.09	0.771	0.041 (0.17%)				

Table 3. Relation between QMUS parameters and time from symptoms onset.

The regression models for thickness were adjusted for sex in biceps/brachialis group, quadriceps femoris, and tibialis anterior and for sex and BMI in forearm flexors. The regression models for echointensity were adjusted for sex and BMI in biceps/brachialis group, and for sex in forearm flexors and tibialis anterior. The regression models for echovariation did not need corrections. SE: Standard Error for coefficient B. 95% CI: 95% Confidence Interval. *Rp: partial correlation coefficient and partial determination coefficient in brackets. Notice that in these analyses, the independent variable was the time from symptoms onset, and the dependent variables were QMUS parameters.

Tuble in Relations between Quiebs parameters and mixe besie.										
QMUS-MRC	_	Raw		Adjusted						
	B coefficient (SE)	95% CI for B	p-value	Rp (Rp-squared %)*	B coefficient (SE)	95% CI for B	p-value	Rp (Rp-squared %)		
Biceps/brachialis	group (Model Goodne	ss = 58.3%								
Thickness	1.38 (0.338)	0.71 to 2.05	< 0.001	0.378 (14.3%)	1.98 (0.347)	1.29 to 2.67	< 0.001	0.498 (24.8%)		
Echointensity	-0.32 (0.186)	-0.69 to 0.05	0.087	-0.17 (2.9%)	-0.34 (0.173)	-0.68 to 0	0.053	-0.193 (3.7%)		
Echovariation	1.67 (0.317)	1.04 to 2.3	< 0.001	0.466 (21.8%)	1.51 (0.297)	0.92 to 2.1	< 0.001	0.454 (20.6%)		
Forearm flexor group (Model Goodness= 48.8%)										
Thickness	0.92 (0.278)	0.37 to 1.47	0.001	0.315 (9.9%)						
Echointensity	-0.25 (0.145)	-0.54 to 0.04	0.087	-0.17 (3.7%)						
Echovariation	2.65 (0.383)	1.89 to 3.41	< 0.001	0.569 (32.4%)						
Quadriceps femoris (Model Goodness= 31.2%)										
Thickness	1.59 (0.295)	1 to 2.17	< 0.001	0.475 (22.5%)						
Echointensity	-0.13 (0.166)	-0.46 to 0.2	0.451	-0.075 (0.6%)						
Echovariation	0.95 (0.466)	0.03 to 1.87	0.044	0.2 (4.0%)						
Tibialis anterior (Model Goodness= 54.9%)										
Thickness	1.15 (0.408)	0.34 to 1.96	0.006	0.272 (7.4%)						
Echointensity	-0.17 (0.132)	-0.43 to 0.09	0.201	-0.128 (1.6%)						
Echovariation	2.49 (0.321)	1.85 to 3.12	< 0.001	0.612 (37.5%)						

Table 4. Relations between QMUS parameters and MRC score.

The regression models were adjusted for sex in biceps/brachialis group. SE: Standard Error for coefficient B. 95% CI: 95% Confidence Interval. *Rp: partial correlation coefficient and partial determination coefficient (Rp-squared) in brackets. The independent variables were the QMUS parameters and the dependent variable was MRC score.

QMUS-ALSFRS		Raw		Adjusted						
	B coefficient (SE)	95% CI for B	p-value	Rp (Rp-squared %)*	B coefficient (SE)	95% CI for B	p-value	Rp (Rp-squared %)		
Biceps/brachialis g	roup (Model Goodne	ess= 54.9%)								
Thickness	0.69 (0.169)	0.35 to 1.02	< 0.001	0.375 (14.1%)	1.03 (0.17)	0.69 to 1.36	< 0.001	0.519 (26.9%)		
Echointensity	-0.1 (0.093)	-0.29 to 0.08	0.270	-0.11 (1.2%)	-0.11 (0.085)	-0.28 to 0.05	0.183	-0.134 (1.8%)		
Echovariation	0.94 (0.159)	0.63 to 1.26	< 0.001	0.51 (26%)	0.85 (0.146)	0.56 to 1.14	< 0.001	0.506 (25.6%)		
Forearm flexor group (Model Goodness= 46.1%)										
Thickness	0.39 (0.138)	0.12 to 0.67	0.005	0.275 (7.5%)						
Echointensity	-0.12 (0.072)	-0.26 to 0.02	0.104	-0.162 (2.6%)						
Echovariation	1.45 (0.19)	1.07 to 1.83	< 0.001	0.606 (36.7%)						
Quadriceps femoris (Model Goodness= 29.9%)										
Thickness	0.73 (0.149)	0.44 to 1.03	< 0.001	0.441 (19.4%)	0.68 (0.151)	0.38 to 0.98	< 0.001	0.414 (17.2%)		
Echointensity	-0.1 (0.084)	-0.26 to 0.07	0.250	-0.115 (1.3%)	-0.12 (0.082)	-0.28 to 0.05	0.156	-0.143 (2%)		
Echovariation	0.67 (0.235)	0.2 to 1.13	0.006	0.273 (7.4%)	0.71 (0.228)	0.25 to 1.16	0.003	0.299 (8.9%)		
Tibialis anterior (Model Goodness= 57	7.4%)								
Thickness	0.56 (0.205)	0.15 to 0.96	0.008	0.263 (6.9%)	0.82 (0.219)	0.39 to 1.26	< 0.001	0.355 (12.6%)		
Echointensity	-0.13 (0.066)	-0.27 to 0	0.045	-0.199 (4.0%)	-0.16 (0.065)	-0.29 to -0.03	0.014	-0.246 (6%)		
Echovariation	1.2 (0.161)	0.88 to 1.52	< 0.001	0.598 (35.8%)	0.98 (0.167)	0.65 to 1.31	< 0.001	0.509 (25.9%)		

Table 5. Relations between QMUS parameters and ALSFRS-r score.

The regression models ALSFRS-r were adjusted for sex in biceps/brachialis group, for sex and BMI in quadriceps femoris and tibialis anterior. SE: Standard Error for coefficient B. 95% CI: 95% Confidence Interval. *Rp: partial correlation coefficient and partial determination coefficient (Rp-squared) in brackets. The independent variables were the QMUS parameters and the dependent variable was ALSFRS-r score

£

Figure 1 Click here to download high resolution image



Figure 2 Click here to download high resolution image

